Ebola Virus Infection: Life Cycle, Prevention, Controland Its Treatment Opportunities

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ABSTRACT

Ebola is a serious and deadly virus transmitted by animals and humans. It is come under the Filoviridae family. It causes very high fever, which lead to bleeding in the body (inside and outside), thereby also called as hemorrhagic fever. Ebola is further classified into subtypes (Bundibugyo, Reston, Sudan, Taï Forest, Zaire) according to location where they were identified. This virus is transmitted by animals (primates only) to human, therefore it is called as zoonotic virus. Human to human transfer is also found in Ebola virus infection. There is no drug is approved by US-FDA till date for this disease.

In this review, we summarized the pathogenesis of infection as well as importance of different proteins in Ebola life cycle. In addition, treatment, control and management of the Ebola disease are also highlighted. Indeed, in recent advances about the structure and biology of Ebola virus and itscharacteristics in the pathogenesis and further treatment opportunities are also covered. Moreover, drug repurposing efforts to identify the novel therapeutic effects/uses of the existing drugs are also discussed. During these studies, it was found that few drugs used for cardiovascular diseases and malaria (FDA approved: Verapamil, Nimodipine and Diltiazem and Chloroquine) have also exhibited potential against Ebola virus infection. Moreover, natural product obtained from Chinese and Japanese herbs (Tetrandrine), have inhibitory effect against Ebola virus. These finding can also be exploited for further drug discovery research.

Keywords: Ebola virus lifecycle, Transmission, Sign and symptoms, Proteins function, Treatment.

Introduction

Ebola virus disease (EVD) sporadic and deadly disease caused by Ebola virus. There are five virus strain has been found according to different location of world in different pathophysiological conditions. Ebola hemorrhagic fever is another name of the EVD. According to the World Health Organization (WHO), there is up to 90% fatality rate in occurrence of EVD. It was first identified in Democratic Republic of the Congo and Sudan in 1976. Since then, there have been intermittentepidemics in Africa. In 1989, the Reston Ebola virus was first discovered (in USA). The monkeys originated from Philippines (crab-eating macaque), were infected from virus and died. In 2009, Reston Ebola virus antibodies were found in the six people infected from pigs in the Philippines with no symptoms. This virus is not highly pathogenic for human as compared to other Ebola viruses. However, WHO has been public health alert to the Asia-Pacific region about the newly emerging disease in animals and humans.

WHO reported that total number of 5,459 deaths were reported in six affected countries (the United States of America Guinea, Spain, Sierra Leone, Mali, andLiberia) and two previously reported countries (Senegaland Nigeria) [WHO report on Ebola response roadmap Situation: 1 October 2014] [1-4]. According to Centers for Diseases Control and Prevention (CDC) (USA), Ebola virus infections (28,639 new cases) and deaths (11,316 cases) were reported in March 2016. All over the world, Ebola virus is mostly found in remote village in Central and West Africa [2, 3, 4].

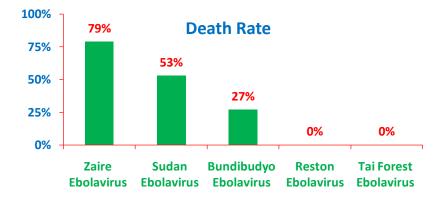


Fig1:Death rates of the 5 Ebola virus species [1-7]

Etiology and Transmission

Ebola is zoonotic virus (spread to the humans from non-human animals). The natural host of this virus is Fruit bats. There are five types of Ebola virus have been foun din different geographical location (See below Table 1).

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Table 1: Ebola virus belongs to the Filoviridae family. It comprises five distinct species [4-5]

Name of Ebola Virus	Geographical location
Bundibudyo Ebolavirus (BDBV)	Uganda
Zaire Ebolavirus (EBOV)	Congo and Gabonese Republic
Sudan Ebolavirus (SUDV)	Sudan and Uganda
Reston Ebolavirus (RESTV)	Philippines
Tai Forest Virus (TAFV)	Republic of Côte d'Ivoire/Ivory Coast

In chimpanzees and gorillas, EBOV and TAFV species were observed in 1994. It has been found that person who handing these infected (Gorillas, Monkeys, Porcupines, Chimpanzees, Fruit bats, Forest antelope) were ill or dead in Africa[4, 5]. Handling of these animal people can get infected with Ebola virus (documented by WHO).

Transmission from animals to humans

If human is having the close contact with body secretion of infected animals (organ or body fluid and blood secretion), then infection is transmitted to the human. Here is the main cause of the transmission [6-8].

- **Blood-**Eating infected/Slaughtering animals
- Waste products-In African caves and underground mine workers (close contact with the urine or feces of infected bats). [7,8]

Transmission from human to human

The human to human transmission is possible through direct contact. It may easily transfer to thousands of people. This endemic very difficult to control. It can be occurred by several ways, including[6-9]:

- Direct contact (Mucus membrane, body secretions/fluids, and broken skin)
- Indirect contact (from the environment if it is contaminated with virus)
- Semen of the infected people and people or who just recovered from EVD

This disease does not contagious until symptoms present in the infected person. Most importantly family members are always on the high risk if one member got infected with the Ebola virus. Medical practitioner or doctor may get infected if they do not use protective care (as per WHO). Every medical or surgical equipment's need to be sterilized properly before again use to other patients [6-7].

Signs and Symptoms

EVDcharacterized by abrupt high fever (not present in generally) is a severe acute viral infection withthe symptoms like: intense headache, sore throat, severe weakness and muscle ache. After some time, this infection is appeared to have other symptoms likerash, vomiting, impaired kidney, diarrhea, and liver function (high level of liver enzymes). In some cases, bleeding may also present in inside and outside the body (according to WHO). According to WHO, symptoms is appeared after 2-21 days of infection. Person is having the virus in semen of the infected man after the 61 days of onset of disease. Now person can able to transmit the disease to other human. The symptoms are extremely nonspecific in the beginning- Ebola looks like almost anything (similar to other disease)[4, 6-9].

According to the CDC, the patient may also have the following symptoms [WHO report, 2015]: Meningitis, Typhoidfever, Malaria, Hepatitis, Shigellosis, Cholera, Relapsing fever, Leptospirosis, Plague, Rickettsiosis, Abdominal pain and Other viral hemorrhagic fevers. At the late stage of disease, bleeding from mouth and nose, this called hemorrhagic syndrome. Typically, this is present in 30 to 50% of patients.

Series of common diagnostic tests are used for diagnosis of EVD as per follow (from WHO Report)[4, 6, 8-10]:

- Serum neutralization test
- RT-PCR (Reverse transcriptase- polymerase chain reaction) assay
- Cell culture for virus isolation
- Antigen-capture detection tests
- Antibody-capture ELISA (enzyme-linked immunosorbent assay)

Life cycle of Ebola virus

Ebola Virus Genome and its proteins

• The EBOV belongs to Filoviridae family and having RNA virus. The 19-kb linearssRNA which is non-segmented. This genome encodes different structural and non-structural proteins. These are nucleoprotein (NP), virion protein 35 (VP35), VP30, 3' non-coding region (leader), VP40, glycoproteins (likesGP, ssGP, GP1,2), VP24, 5' non-coding region and RNA dependent RNA-polymerase[11].

Functions of different proteins in Ebola life cycle:

- GP protein is responsible for important functions in extracellular environment and the virion proteins (VP40, VP35 and VP24) are having intracellular roles (Details is given in **Table 2**).
- The GP gene encode for three different products which consist of soluble GP (sGP) [lacks of transmembrane domain] and small soluble GP (ssGP), GP1 (receptor binding) and GP2 (viral fusion) subunits[12, 13].
- GP is located on the virion surface (only viral protein). This is exerting very important functions like attachment and fusion of the virion to the host [14-16].
- By GP1,2 is synthesized by transcriptional RNA editing. After the post-translation, it is cleaved into two disulphide subnits; one is surface unit which is called GP1 and other is membrane unit which is called GP2 [17-18].
- GP1 contains receptor binding site (RBS) to attach to host cell and a domain which defend the RBD from immune systems. For viral entry (into host), receptor binding and cellular tropism, RBS of GP1 is responsible for it [16, 17].
- GP2 helps in binding of membrane of host endosomal cell (acidic pH) to the viral membrane (through macropinocytosis). Acidic pH of the endosomes is responsible for cleavages of the GP1 [16-18].
- Acidic environment is present in endosomes. It is important for activation of cysteine proteases:Cathepsin B and Cathepsin L. These proteases are responsible for the cleavage of EBOV.
- The fusion between endosomal and virusmembrane is done by GP1,2 [16-17]. After of the virus entry, two receptors are found to important for fusion that are the Niemann-Pick C1 (NPC1) receptor and two-pore segment channel 2 (TPC2). At this time, the conformational changes occurred in the GP2. These areenabling the release of viral genome into the cytoplasm [16, 19-20].
- Host immune system is suppressed by VP35 and VP24 proteins.
- Using host system, Ebola virus is synthesizing the new virions via transcription and replication of RNA with help of VP35, VP30, and NP, L-protein.VP40 and VP24 is having role in replication and transcription regulation. However, VP30 is responsible for transcription initiation only. These available protein in Ebola virus can be used as an attractive target for drug design and discovery for new therapeutic intervention [16-20].
- For virus assembly and budding, matrix protein VP40 is very much important [16, 18, 20].

Table2: Functions of different Ebola proteins in its life cycle [21]

Viral	Main Functions	Other Functions
Protein	- 1- 1-1-1 -1	3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3
L	-Regulation transcription	-
VP40	-Viral budding and assembly	-HampersRNA interference
NP	-Protection of viral genome from	-
	degradation by encapsulation	
	-Important part of RNA protein complex	
VP30	-Initiates transcription	Impedes RNA interference
GP	-Fusion and entry of Ebola virusin host cell	-Reductionsof barrier in endothelial
		cells.
		-Responsible for destruction of T-
		lymphocytes
		-Producing cytotoxicity in cells in later
		stage
VP35	-Inhibits interferon signalling	-Responsible for formation of viral
	-Dendritic cell impairment in maturation	complex with VP30 and NP
	-Impedes RNA interference	-Regulation of RNA synthesis mediated
	-Blocking protein kinase R Inhibits	by NP-RNA interactions
	antiviral effects by	
VP24	-Inhibition ofinterferon signalling by	-Nucleocapsid assembly and stability
	interference of NF-κB and STAT	-Regulation of central dogma process
	signalling pathways	

Different treatment available for EVD

According to Food and Drug Administration (US-FDA), till date there is no drug is approved againstEbola virus. Most of the drugs are in clincal trial (soon may available for the treatment) (Source: https://www.fda.gov/emergency-preparedness-and-response/mcm-issues/ebola-preparedness-and-response-updates-fda, accessed on date: 27-11-2019).But some guideline is given by WHO for incentive supportive care in Ebola virus infection, and often includes:

- Patient's fluids and electrolytes balancing
- Keepingthe blood pressure and oxygen status normal
- For any complicated infection, treatment has to take by a patient

Several vaccines are designed and tested [found satisfactory in animal model], but not available for clinical use [6, 7-9].Recently, a breakthrough for the treatment of Ebola virus was announced in August 2019 using monoclonal antibodies. The monoclonal antibodies (REGN-EB3 and mAb114) are found very good response in randomized controlled trial. It was found that the single dose REGN-EB3 intravenous treatment is decreased the mortality in the patients, especially in early stage strongly. On other hand mAb114 is having the similar effect as REGN-EB3. The US-FDA have been given orphan drug status (licensing not done)

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[11, 22]. Both are being made available to all patients in the Congo. Moreover, first human clinical trials data are become available for the following three candidate vaccines (See below Table):

Name of the Vaccine	Developed by	Dose	References
rVSV-ZEBOV (V920)	Merck	Single dose	[23]
Ad26-ZEBOV/MVA-	Bavarian Nordic	Prime-boost regimen	[24]
BN-Filo	GmbH		
ChAd3-EBO-Z	Bavarian Nordic GmbH	Tested in either a single dose (GlaxoSmithKline) or in combination with a booster dose of other vaccine (MVA-BN-Filo vaccine)	[25]

Some of the drugs and vaccines are also found in the literatures which is found promising effect in Ebola virus infection. However, lot of Ebola inhibitors (tested in vitro and in vivo studies) have also been listed in the literature(see **Table 3**) [26].

Table 3: Ebola therapeutics reported in different research studies [18]

Chemical	Action	Company	US-FDA Status	Refere
Agent		Status		nces
T-705	Broad-spectrum antiviral drug:	Toyama	Not approved	[27-29]
(Favipiravir)	against many RNA virus	Chemical	(found	
		of Japan	promising)	
BCV,	Active against dsDNA viruses,	Chimerix	Not approved	[30-31]
CMX001	however Ebola is dsRNA virus	of Durham,	(found	
(Brincidofovir)		NC,	promising)	
BCX4430;	Broad-spectrum antiviral	BioCryst	No plan for	[32]
Immucillin-A	effectiveness against a range of	Pharmaceu	Ebola virus	
(Galidesivir)	other RNA virus	ticals	testing	
JK-05	Inhibitor of the viral enzyme	Sihuan	Not approved	[18]
	RNA polymerase	Pharmaceu	(Copy of	
		tical	Favipiravir)	
TKM-Ebola	This is small interfering RNAs.	Arbutus	Phase II trial	[33]
	It is targeting three proteins in	Biopharma	failed, so	
	Ebola virus life cylce: VP24,	(formerly	changed to	
	VP35, and Zaire Ebola L	Tekmira	hepatitis B	
	polymerase	Pharmaceu	testing	
		ticals		
		Corp.)		
ZMapp	Ebola glycoprotein epitopes	license by	Not approved yet	[34]

(monoclonal antibodies)		Mapp Biopharma -ceutical	(in clinical trial)	
NSC62914	Scavenger of reactive oxygen species		Not approved (because of compound toxicity)	[35-36]
FGI-103	Replication inhibitor (not yet completely)	-	Not approved	[37]
FGI-104	Inhibiting the protein TSG101	-	Not approved	[38-39]
FGI-106	Not complete known, but speculated viral entry inhibitor active against positive or negative RNA virus	-	Not approved	[18, 26, 40]

Some of the drugs that emerged from already available marketed drug by drug repurposing approach. This means, drug is invented for one disease but also found new therapeutic use in other disease. Some of the drugs which identified through this approach are mentioned in the **Table4**.

Table4: Available drugs which are identified by repurposing process

Name of Drugs	Repurposed of the drug in Ebola	Parent function	Reference
	infection		
Convalescent blood serum	 Neutralizing antibodies and T-cell responses IgG level increased VP40 and NP directly inhibited Clearing the viral antigen from the circulation and cytotoxic T cells activation 	Whole blood or plasma from a patient who has survived a previous infection, considered to be especially rich in antibodies against the infectious agent of the disease (WHO report)	[41-42]
Chloroquine	 Able to increase endosomal pH As an entry inhibitor for many viruses Targeting the Cathepsins L cleavage phenomena (A broad-spectrum) 	Anti-malarial	[43]
Cationic amphiphiles	Amiodarone: Potent entry	Used in	[44-49]

(Dronedarone, Amiodarone, Verapamil, Clomiphene, and Toremifene)	 inhibitor (Niemann-Pick type-C1 dependent). Clomiphene, and Toremifene: Entry inhibitor (indirectly through NPC1) Dronedarone: Similar to amiodarone activity Verapamil: Similar to amiodarone activity 	cardiovascular diseases	
Na+/K+ exchangers (amiloride and its derivatives)	Potent dose-dependent (at non-cytotoxic dose) as an entry inhibitor and inhibition of infection of the EBOV.	Diuretics (Potassium sparing) to treat hypertension and cardiovascular disease	[50]
Na+/K+-ATPase pump inhibitors (Cardiac glycosides)	 Ouabain: inhibitthe EBOV replication (in human MRC-5 cells at non-cytotoxic conc.). Digoxin, and Digitoxin: Role is not clearly understood. 	This are cardiac glycosides (used in cardiovascular disease)	[51-55]

Prevention and control

According WHO, these are the precautions that can be used to reduce the chance of infection and control [42].

Controlling in domestic animals

- Regularlywashingof the animal (Pig or monkey farms).
- Disinfection of animals with NaOCl (Sodium Hypochlorite) or other detergents.
- If virus infection is suspected, then immediately premises should be closed for no use further or quarantined.
- Close supervision of infected animal and its isolation from the non-infected animals.
- Isolate the infected animal from the other animal and make restricted movement of animals to other non-infected areas. This can reduce the spread of the disease.
- Safely incineration or burial of corpses (dead infected animals).
- Animal health surveillance system should be established and new case must be informed early to the human public healthand veterinaryauthorities[1, 4, 7, 8].

Reducing the risk of Ebola infection in people

- Gloves or other protective items must be used for handling of the infected Animals and people.
- Cook the food [Animal products (blood and meat)] thoroughly before take.

- Regular cleaning of the hospitals and medical equipment's should be done from the disinfectants after visiting the infected person.
- Safely buried of the infected people [1, 5, 7-9].

Controlling infection in health-care settings

Health professionals should follow standard safeguards consistently with all patients – regardless of their diagnosisall the time during practices, including [4, 6-10]:

- Respiratory hygiene
- Safe burial practices
- Use of personal protective equipment (according to contact with infected materials)
- Safe injection practices
- Basic hand hygiene

Risk factors for Ebola

Person can be on high risk if:

- Traveling to areas of Africa (where infected people is living or confirmed case of Ebola is present).
- From Africa or the Philippines, importing animals for research work
- Involve in the medical care of the infected people and protection must be taken to care the infected people like sterilized syringes or cotton, use gloves and mask, etc.
- During cremation of dead body (infected people). [1, 4, 7-9, 11]

Conclusions

There is an alarm for EVD as it is contagious and very less opportunities for the treatment in the market. It is mainly found in West Africa, Sudan, Congo and. But it is sporadically found in other country due to tourist visited in the above-mentioned countries. The WHO gave few guidelines for prevention, control and treatment of the disease. But still a challenge for control of this disease as it is transmitted through the human to human interactions also. However, 90% transmission occurs through the infected animals.

In the life cycle of Ebola virus lot of proteins [NP, VP35, VP30, VP40, VP24, glycoproteins (like sGP, ssGP, GP1,2) and RNA dependent RNA-polymerase] are present which are pivotal role in multiplication and survival of the virus in the human. Infected people get immunocompromised due to attack of the virus in the human immune system. These proteins can be used to design and discovery novel drugs.

As of now, there is no reported treatment for the virus. However, some of the drugs are being used for the treatment of the Ebola infection. It was reported that few marketed drugs (used in

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cardiovascular disease) can be used Ebola virus infections/diseases, like Verapamil, Nimodipine and Diltiazem. In addition, antimalarial drug (Chloroquine) is found to have effect in Ebola Disease. Moreover, Chinese and Japanese herbs Tetrandrine, have inhibitory action on Ebola virus. These drugs can be used further designing/identification of novel therapeutics. These finding suggested that Ebola is having different drug targets as also present in other virus, which can be further exploited to design and discovery novel drugs/leads with excellent potency and efficacy.

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References

- 1. World Health Organization. Clinical management of patients with viral haemorrhagic fever: A pocket guide for the front-line health worker. http://apps.who.int/iris/bitstream/10665/130883/2/WHO_HSE_PED_AIP_14.05.pdf?ua=1 (Accessed on 27, Nov. 2019).
- 2. WHO Ebola Response Team, "Ebola virus disease in West Africa—the first 9 months of the epidemic and forward projections", N. Engl. J. Med., Vol. 371, No. 16, pp. 1481-1495, Oct. 2014.
- 3. Mike Bray, and Frederick A. Murphy, "Filovirus research: knowledge expands to meet a growing threat", The J. Infect. Dis., Vol. 196, No. Suppl._2, pp. S438-S443, Nov. 2007.
- 4. EmanueleNicastri, Gary Kobinger, Francesco Vairo, Chiara Montaldo, Leonard EG Mboera, Rashid Ansunama, AlimuddinZumla, and Giuseppe Ippolito, "Ebola Virus Disease: Epidemiology, Clinical Features, Management, and Prevention", Infect. Dis. Clin. North Am., Vol. 33, No. 4, pp.953-976, Nov. 2019.
- 5. Emanuel, Jackson, Andrea Marzi, and Heinz Feldmann. "Filoviruses: ecology, molecular biology, and evolution." In Advances in Virus Research, Vol. 100, pp. 189-221. Academic Press, 2018.
- 6. HeinzFeldmann, and Thomas W. Geisbert, "Ebola haemorrhagic fever", The Lancet, Vol. 377, No. 9768, pp. 849-862, Mar. 2011
- 7. DiegoCantoni, Arran Hamlet, Martin Michaelis, Mark N. Wass, and Jeremy S. Rossman, "Risks posed by Reston, the forgotten Ebolavirus." mSphere, Vol. 1, No. 6, pp. e00322-16, 2016.
- 8. Al-Tameemi, K. A. N. A. A. N., and RaiaanKabakli, "Ebola Virus: An Overview Disease and Treatment", Asian J. Pharmaceut. Clinc. Res., Vol 12, No. 10, pp. 57-62, Oct. 2019.
- 9. Baseler, Laura, Daniel S. Chertow, Karl M. Johnson, Heinz Feldmann, and David M. Morens, "The pathogenesis of Ebola virus disease", Annu. Rev. Pathol.-Mech., Vol. 12, pp. 387-418, Jan. 2017.
- 10. Negredo, Ana, Gustavo Palacios, Sonia Vázquez-Morón, Félix González, HernánDopazo, Francisca Molero, Javier Juste et al., "Discovery of an ebolavirus-like filovirus in europe", PLoSPathog., Vol. 7, No. 10, pp. e1002304, Oct. 2011.
- 11. Thomas Hoenen, Allison Groseth, Darryl Falzarano, and Heinz Feldmann, "Ebola virus: unravelling pathogenesis to combat a deadly disease", Trends Mol. Med., Vol. 12, No. 5, pp. 206-215, May 2006.

Page | 3854 Copyright ⊚ 2019Authors

- 12. Valentina A. Volchkova, Heinz Feldmann, Hans-Dieter Klenk, and Viktor E. Volchkov, "The nonstructural small glycoprotein sGP of Ebola virus is secreted as an antiparallel-orientated homodimer", Virol., Vol. 250, No. 2, pp. 408-414, Oct. 1998.
- 13. Masfique Mehedi, Darryl Falzarano, Jochen Seebach, Xiaojie Hu, Michael S. Carpenter, Hans-Joachim Schnittler, and Heinz Feldmann, "A new Ebola virus nonstructural glycoprotein expressed through RNA editing", J. Virol., Vol. 85, No. 11, pp. 5406-5414, Mar. 2011.
- 14. Masayuki Shimojima, Ayato Takada, Hideki Ebihara, Gabriele Neumann, Kouki Fujioka, TatsuroIrimura, Steven Jones, Heinz Feldmann, and Yoshihiro Kawaoka, "Tyro3 family-mediated cell entry of Ebola and Marburg viruses", J. Virol., Vol. 80, No. 20, pp. 10109-10116, 2006.
- 15. Jeffrey E. Lee, and Erica Ollmann Saphire, "Ebolavirus glycoprotein structure and mechanism of entry", Future Virol. 4, No. 6, pp. 621-635, Nov. 2009.
- 16. Sven Moller-Tank and Wendy Maury, "Ebola virus entry: a curious and complex series of events", PLoSPathog., Vol. 11, No. 4, pp. e1004731, Apr. 2015.
- 17. Kang Yiu Lai, Wing Yiu George Ng, and Fan Fanny Cheng, "Human Ebola virus infection in West Africa: a review of available therapeutic agents that target different steps of the life cycle of Ebola virus", Infect. Dis. Poverty, Vol. 3, No. 1, pp. 43, Nov. 2014.
- 18. Paul E. Kilgore, John D. Grabenstein, Abdulbaset M. Salim, and Michael Rybak, "Review of Therapeutics", Pharmacotherapy, Vol. 35, No. 1, pp. 43-53, 2015.
- 19. Rebecca M. Mingo, James A. Simmons, Charles J. Shoemaker, Elizabeth A. Nelson, Kathryn L. Schornberg, Ryan S. D'Souza, James E. Casanova, and Judith M. White, "Ebola virus and severe acute respiratory syndrome coronavirus display late cell entry kinetics: evidence that transport to NPC1+ endolysosomes is a rate-defining step", J. Virol., Vol. 89, No. 5, pp. 2931-2943, Dec. 2015.
- 20. James A. Simmons, Ryan S. D'Souza, Margarida Ruas, Antony Galione, James E. Casanova, and Judith M. White, "Ebolavirus glycoprotein directs fusion through NPC1+ endolysosomes", J. Virol., Vol. 90, No. 1, pp. 605-610, Oct. 2016.
- 21. Diego Cantoni, and Jeremy S. Rossman, "Ebolaviruses: New roles for old proteins", PLoSNegl. Trop. Dis., Vol. 12, No. 5, pp. e0006349, May 2018
- 22. Daniel R Lucey, "New treatments for Ebola virus disease", Br. Med. J., Vol. 366, pp. 5371, Sept. 2019.
- 23. Ana MariaHenao-Restrepo, Anton Camacho, Ira M. Longini, Conall H. Watson, W. John Edmunds, Matthias Egger, Miles W. Carroll et al., "Efficacy and effectiveness of an rVSV-vectored vaccine in preventing Ebola virus disease: final results from the Guinea ring vaccination, open-label, cluster-randomised trial (Ebola ÇaSuffit!)", The Lancet, Vol. 389, No. 10068, pp. 505-518, Feb. 2017.
- 24. Zacchaeus Anywaine, Hilary Whitworth, Pontiano Kaleebu, George Praygod, Georgi Shukarev, Daniela Manno, Saidi Kapiga, Heiner Grosskurth, Samuel Kalluvya, Viki Bockstal, Dickson Anumendem, Kerstin Luhn, Cynthia Robinson, Macaya Douoguih and Deborah Watson-Jones, "Safety and Immunogenicity of a 2-Dose Heterologous Vaccination Regimen With Ad26. ZEBOV and MVA-BN-Filo Ebola Vaccines: 12-Month Data From a Phase 1 Randomized Clinical Trial in Uganda and Tanzania", The J. Infect. Dis., Vol. 220, No. 1, pp. 46-56, Jun. 2019.
- 25. Tapia, Milagritos D., Samba O. Sow, Kirsten E. Lyke, Fadima Cheick Haidara, Fatoumata Diallo, Moussa Doumbia, Awa Traore et al., "Use of ChAd3-EBO-Z Ebola virus vaccine in Malian and US adults, and boosting of Malian adults with MVA-BN-Filo: a phase 1, single-blind, randomised trial, a phase 1b, open-label and double-blind, dose-escalation trial, and a nested, randomised, double-blind, placebo-controlled trial", Lancet Infect. Dis., Vol. 16, No. 1, pp. 31-42, Jan. 2016.

Page | 3855 Copyright ⊚ 2019Authors

- 26. EdwigePicazo, and Fabrizio Giordanetto, "Small molecule inhibitors of ebola virus infection", Drug Discov. Today, Vol. 20, No. 2, pp. 277-286, Feb. 2015.
- 27. LisaOestereich, Anja Lüdtke, Stephanie Wurr, Toni Rieger, César Muñoz-Fontela, and Stephan Günther, "Successful treatment of advanced Ebola virus infection with T-705 (favipiravir) in a small animal model", Antiviral Res., Vol. 105, pp. 17-21, May 2014.
- 28. JeremieGuedj, Geraldine Piorkowski, Frederic Jacquot, Vincent Madelain, ThiHuyen Tram Nguyen, Anne Rodallec, Stephan Gunther, Caroline Carbonnelle, France Mentre, Herve Raoul, Xavier de Lamballerie, "Antiviral efficacy of favipiravir against Ebola virus: A translational study in cynomolgus macaques", PLoS Med., Vol. 15, pp. e1002535, Mar. 2018.
- 29. YousukeFuruta, Takashi Komeno, and TakaakiNakamura, "Favipiravir (T-705), a broad-spectrum inhibitor of viral RNA polymerase", Proc. Jpn. Acad. Ser. B Phys. Biol. Sci., Vol. 93, No. 7, pp. 449-463, Aug. 2017.
- 30. Debra C. Quenelle, Bernhardt Lampert, Deborah J. Collins, Terri L. Rice, George R. Painter, and Earl R. Kern, "Efficacy of CMX001 against herpes simplex virus infections in mice and correlations with drug distribution studies", The J. Infect. Dis., Vol. 202, No. 10, pp. 1492-1499, Nov. 2010.
- 31. Diana F.Florescu, and Megan A. Keck, "Development of CMX001 (Brincidofovir) for the treatment of serious diseases or conditions caused by dsDNA viruses", Expert Rev. Anti Infect. Ther., Vol. 12, No. 10, pp. 1171-1178, Oct. 2014.
- 32. Raymond Taylor, Pravin Kotian, Travis Warren, Rekha Panchal, SinaBavari, Justin Julander, Sylvia Dobo, Angela Rose Yahya El-Kattan Brian Taubenheim, Yarlagadda Babu William P.Sheridan, "BCX4430—a broad-spectrum antiviral adenosine nucleoside analog under development for the treatment of Ebola virus disease", J. Infect, Public Health, Vol. 9, No. 3, pp. 220-226, May 2016.
- 33. Colleen S. Kraft, Angela L. Hewlett, Scott Koepsell, Anne M. Winkler, Christopher J. Kratochvil, LuAnn Larson, Jay B. Varkey et al., "The use of TKM-100802 and convalescent plasma in 2 patients with Ebola virus disease in the United States", Clin. Infect. Dis., Vol. 61, No. 4, pp. 496-502, Aug. 2015.
- 34. Madelain, Vincent, ThiHuyen Tram Nguyen, AnaelleOlivo, Xavier De Lamballerie, JeremieGuedj, Anne-Marie Taburet, and France Mentré, "Ebola virus infection: review of the pharmacokinetic and pharmacodynamic properties of drugs considered for testing in human efficacy trials", Clin. Pharmacokinet., Vol. 55, No. 8, pp. 907-923, Aug. 2016.
- 35. Marina Jerebtsovaand Sergei Nekhai, "Therapeutics for postexposure treatment of Ebola virus infection", Future Virol., Vol. 10, No. 3, pp. 221-232, Mar. 2015.
- 36. Rekha G. Panchal, Julie P. Tran, Alison A. Bergeron, Jay Wells, Krishna P. Kota, Javad Aman, and SinaBavari, "Identification of an antioxidant small-molecule with broad-spectrum antiviral activity", Antiviral Res., Vol. 93, No. 1, pp. 23-29, Jan. 2012.
- 37. Travis K. Warren, Kelly L. Warfield, Jay Wells, Sven Enterlein, Mark Smith, Gordon Ruthel, Abdul S. Yunus, Michael S. Kinch, Michael Goldblatt, M. Javad Aman and SinaBavari, "Antiviral activity of a small-molecule inhibitor of filovirus infection", Antimicrob. Agents Chemother., Vol. 54, No. 5, pp. 2152-2159, Mar. 2010.
- 38. Michael S. Kinch, Abdul S. Yunus, Calli Lear, Hanwen Mao, Hanson Chen, Zena Fesseha, Guangxiang Luo Eric A. Nelson, Limin Li, Zhuhui Huang, Michael Murray, William Y. Ellis, Lisa Hensley, Jane Christopher-Hennings, Gene G. Olinger, and Michael Goldblatt, "FGI-104: a broad-spectrum small molecule inhibitor of viral infection", Am. J. Transl. Res., Vol. 1, No. 1, pp. 87-98, Jan. 2009.
- 39. ZlatkoJaneba, "Development of Small-Molecule Antivirals for Ebola", Med. Res. Rev., Vol. 35, No. 6, pp. 1175-1194, Nov. 2015.

- 40. M. Javad Aman, Michael S. Kinch, Kelly Warfield, Travis Warren, Abdul Yunus, Sven Enterlein, Eric Stavale, Peifang Wang, Shaojing Chang, Qingsong Tang, Kevin Porter, Michael Goldblatt and SinaBavari, "Development of a broad-spectrum antiviral with activity against Ebola virus", Antiviral Res., Vol. 83, No. 3, pp. 245-251, Sept. 2009.
- 41. GiuseppeMarano, Stefania Vaglio, Simonetta Pupella, Giuseppina Facco, Liviana Catalano, Giancarlo M. Liumbruno, and Giuliano Grazzini", Convalescent plasma: new evidence for an old therapeutic tool?", Blood Transfus., Vol. 14, No. 2, pp. 152-157, Mar. 2016
- 42. https://www.who.int/csr/resources/publications/ebola/convalescent-treatment/en/
- 43. Peter B. Madrid, Sidharth Chopra, Ian D. Manger, Lynne Gilfillan, Tiffany R. Keepers, Amy C. Shurtleff, Carol E. Green, Lalitha V. Iyer, Holli Hutcheson Dilks, Robert A. Davey, Andrey A. Kolokoltsov, Ricardo Carrion Jr, Jean L. Patterson, SinaBavari, Rekha G. Panchal, Travis K. Warren, Jay B. Wells, Walter H. Moos, RaeLyn L. Burke, Mary J. Tanga, "A systematic screen of FDA-approved drugs for inhibitors of biological threat agents", PloSOne, Vol. 8, No. 4, pp. e60579, Apr. 2013.
- 44. Gerrit Gehring, Katrin Rohrmann, NkachehAtenchong, Eva Mittler, Stephan Becker, Franziska Dahlmann, Stefan Pöhlmann, Florian W. R. Vondran, Sascha David, Michael P. Manns, Sandra Ciesek, Thomas von Hahn, "The clinically approved drugs amiodarone, dronedarone and verapamil inhibit filovirus cell entry", J. Antimicrob. Chemother., Vol. 69, No. 8, pp. 2123-2131, Apr. 2014.
- 45. Hatem A.Elshabrawy, Jilao Fan, Christine S. Haddad, KiiraRatia, Christopher C. Broder, Michael Caffrey, and Bellur S. Prabhakar, "Identification of a broad-spectrum antiviral small molecule against severe acute respiratory syndrome coronavirus and Ebola, Hendra, and Nipah viruses by using a novel high-throughput screening assay", J. Virol., Vol. 88, No. 8, pp. 4353-4365, 2014.
- 46. Elizabeth Nelson, Alyson Barnes, Ronald Wiehle, Gregory Fontenot, Thomas Hoenen, and Judith White. "Clomiphene and its isomers block Ebola virus particle entry and infection with similar potency: potential therapeutic implications", Viruses, Vol. 8, No. 8, pp. 206, Aug. 2016.
- 47. Yuguang Zhao, Jingshan Ren, Karl Harlos, Daniel M. Jones, Antra Zeltina, Thomas A. Bowden, Sergi Padilla-Parra, Elizabeth E. Fry, and David I. Stuart, "Toremifene interacts with and destabilizes the Ebola virus glycoprotein", Nature, Vol. 535, No. 7610, pp. 169, Jun. 2016.
- 48. Lisa M. Johansen, Jennifer M. Brannan, Sue E. Delos, Charles J. Shoemaker, Andrea Stossel, Calli Lear, Benjamin G. Hoffstrom, Lisa Evans DeWald, Kathryn L. Schornberg, Corinne Scully, Joseph Lehár, Lisa E. Hensley, Judith M. White and Gene G. Olinger, "FDA-approved selective estrogen receptor modulators inhibit Ebola virus infection", Sci. Transl. Med., Vol. 5, No. 190, pp. 190ra79, Jun. 2013.
- 49. Mohammad F. Saeed, Andrey A. Kolokoltsov, Thomas Albrecht, and Robert A. Davey, "Cellular entry of ebola virus involves uptake by a macropinocytosis-like mechanism and subsequent trafficking through early and late endosomes", PLoSPathog., Vol. 6, No. 9, pp. e1001110, Sept. 2010.
- 50. Isabel García-Dorival, Weining Wu, Stuart Dowall, Stuart Armstrong, Olivier Touzelet, Jonathan Wastling, John N. Barr, David Matthews, Miles Carroll, Roger Hewson, Julian A. Hiscox"Elucidation of the Ebola virus VP24 cellular interactome and disruption of virus biology through targeted inhibition of host-cell protein function", J. Proteome Res., Vol. 13, No. 11, pp. 5120-5135, Aug. 2014.
- 51. Sushma A. Ogram, Christopher D. Boone, Robert McKenna, and James B. Flanegan, "Amiloride inhibits the initiation of Coxsackievirus and poliovirus RNA replication by inhibiting VPguridylylation", Virol., Vol. 464, pp. 87-97, Sept. 2014.

THINK INDIA JOURNAL

ISSN: 0971-1260 Vol-22-Issue-16-August-2019

- 52. Eric Ka-Wai Hui and Debi P. Nayak, "Role of ATP in influenza virus budding", Virol., Vol. 290, No. 2, pp. 329-341, Nov. 2001.
- 53. Luciano Amarelle, Emilia Lecuona, and Jacob I. Sznajder, "Anti-influenza treatment: drugs currently used and under development", Arch. Bronconeumol., Vol. 53, No. 1, pp. 19-26, Jan. 2017.
- 54. H-Heinrich Hoffmann, Peter Palese, and Megan L. Shaw, "Modulation of influenza virus replication by alteration of sodium ion transport and protein kinase C activity", Antiviral Res., Vol. 80, No. 2, pp. 124-134, Nov. 2008.
- 55. Luciano Amarelle, Jeremy Katzen, Masahiko Shigemura, Lynn C. Welch, Héctor Cajigas, Christin Peteranderl, Diego Celli, Susanne Herold, Emilia Lecuona, and Jacob I. Sznajder, "Cardiac glycosides decrease influenza virus replication by inhibiting cell protein translational machinery", Am. J. Physiol.-Lung. Cell. Mol. Physiol., Vol. 316, No. 6, pp. L1094-L1106, May 2019.

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